



RESEARCH

Open Access

Type II and type IV topoisomerase mutations in clinical isolates of *Morganella morganii* harbouring the *qnrD* gene

Mariam Nasri Yaiche^{1,2*}, Ikbel Denden Rafraf², Qinglan Guo³, Maha Mastouri^{2,4}, Mahjoub Aouni² and Minggui Wang³

Abstract

Introduction: The aim of this study was to show the emergence of the *qnrD* gene among fluoroquinolone-resistant *Morganella morganii* isolate. The occurrence of mutations in DNA gyrase (*gyrA* and *gyrB*) and topoisomerase IV (*parC*, *parE*) genes was also investigated in this strain.

Methodology: 95 clinical Enterobacteria were screened for harbouring the *qnrD* gene. The clinical isolate of *M. morganii* was recovered from urine from a patient hospitalized in the urology unit at Fattouma Bourguiba Hospital, Tunisia. Antibiotic susceptibility was tested with the agar disk diffusion method. Quinolone susceptibility was studied with microbroth dilution technique. The investigations of plasmid mediated quinolone resistance (PMQR) and topoisomerases mutations were performed by polymerase chain reaction and nucleotide sequencing.

Results: This isolate showed high level of resistance to quinolones. The MIC with microbroth dilution technique was 512 µg/ml for norfloxacin, 256 µg/ml for ofloxacin and ciprofloxacin and 64µg/ml for levofloxacin. This strain was found to harbour the quinolone resistance determinant *qnrD*. In addition, this strain harboured two new *gyrB* mutations (S463A, S464Y) and one *parC* mutation (S80I).

Conclusions: This is the first report in Tunisia of *qnrD* determinant and two new *gyrB* mutations in *M. morganii*. The nosocomial infection due to this *proteaeae* invites further study of its epidemiologic evolution.

Keywords: *Morganella morganii*, Quinolone resistance, Topoisomerase mutation, *qnrD*

Introduction

The first *qnr* gene was observed in the late 1990s and described as a plasmid-mediated quinolone resistance gene (PMQR). Since that, five types of *qnr* genes have been reported: *qnrA*, *qnrB*, *qnrC*, *qnrD*, and *qnrS*. Their sequences are deposited at the following website (<http://www.lahey.org/qnrStudies/>). Most were observed in *Enterobacteriaceae*, located on large conjugative multi-resistant plasmids, such as pMG252 seen in the original *qnrA1*-positive strain [1].

The *qnrD* gene was first described in 2009 on a 4270 bp plasmid present in a human clinical isolate of

Salmonella enterica serovar *Kentucky* and three *Salmonella enterica* serovar *Bovismordificans* isolates from China [2]. Since this report, *qnrD* was found in *Escherichia coli* [3], *M. morganii* [4], *P. Mirabilis* [4,5], *Pseudomonas aeruginosa* [5] and *Providencia rettgeri* [6].

We describe the new *gyrB* and *parC* mutations in a clinical isolate of *M. morganii* harbouring the rare *qnrD* gene that is first reported in Tunisia.

Material and methods

Case presentation

On 22 November 2012, a 76-years old male presented to the urology department with hematuria and burning micturition that were neglected by the patient since six months. On presentation, he was not febrile with normal vital signs and negative urine culture.

* Correspondence: mariem_nasri@hotmail.com

¹Faculty of Sciences, Bizerta Carthage University, Bizerta, Tunisia

²Laboratory of Biologically Active Substances and Communicable Diseases (LR99ES27), Monastir University, Monastir, Tunisia

Full list of author information is available at the end of the article

The ultrasonography shows a bladder of budding aspect, bifocal with long axis (3 cm). The histopathology revealed urothelial carcinoma infiltrating high-grade bladder (II pTa). On 30/11/2012, underwent a transurethral resection of bladder tumor. A double-current probe 18 c was also introduced.

On 19/01/2013 the patient presented an acute pyelonephritis and the urine culture showed the presence of quinolone resistant *M. morgani*. The empiric antibiotic therapy consisted of three doses a day of 2_g Ampicillin during 4 days. Urine culture subsequently yielded *M. morgani*. The antibiotic regime was then replaced with three doses a day of cefoxitin 1_g during one week. After this, the urine culture was negative but the haematuria still existed.

Bacterial strain

M. morgani was isolated from urine and identified using the Vitek 2 identification system (bioMérieux, Marcy l'Etoile, France). The antimicrobial susceptibilities of the *M. morgani* isolate were determined on Mueller-Hinton agar by the standard disk diffusion procedure as described by the Antibiogram Committee of the French Society for Microbiology (<http://www.sfm.asso.fr>). The microbroth dilution technique was used to determine the minimal inhibitory concentrations (MIC) for norfloxacin, ofloxacin, ciprofloxacin and levofloxacin.

PMQR screening

The *qnrA*, *qnrB*, *qnrS*, *qnrC*, *qnrD*, *aac-Ib*, and *qepA* genes were detected by PCR with the primer sets as described previously [2,7-10].

QnrD primers were used also for the clinical survey of *qnrD* gene in other quinolone resistance *Enterobacteriaceae* (n = 95) collected from October 2010 to June 2013: *Klebsiella pneumoniae* (n = 59), *Enterobacter cloacae* (n = 17), *Citrobacter freundii* (n = 9), *Salmonella spp* (n = 6), *Providencia rettgerii* (n = 3), *M. morgani* (n = 1).

Conjugation

The transferability of *qnrD* gene between the *M. morgani* isolate and the *E. coli* J53Az^F recipient were carried out in LB broth. Cultures of donor and recipient cells in logarithmic phase (0.5 ml of each) were added to 4 ml of fresh LB broth and incubated overnight without shaking. Transconjugants were selected on Trypticase soy agar (TSA) plates containing sodium azide (100 µg/ml; Sigma Chemical Co., St. Louis, Mo.) for counterselection and gentamicin (10 µg/ml) to select for plasmid-encoded resistance.

Detection of mutations in the topoisomerase II and IV genes

The quinolone resistance-determinant regions (QRDRs) of *gyrA*, *gyrB*, *ParC* and *parE* was amplified as described previously [11]. Products resulting from amplifications

were subjected to sequencing. Analysis and comparison of nucleotide and amino acid sequence data were carried out using Lasergene software (version 7.1; DNASTAR, Wisconsin, USA), and programs available from the national Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov>).

Results

Antibiotic susceptibility of *M. morgani*

The clinical strain of *M. morgani* showed a low level of resistance to beta-lactams due to its resistance to ampicillin, amoxicillin, ticarcillin and piperacillin only. It was susceptible to amoxicillin/clavulanic acid, ticarcillin/clavulanic acid, piperacillin/tazobactam, cafalotin, cefamandol, cefoxitin, cefotaxim, ceftazidim, cefpirom, aztreonam, imipenem and ertapenem. This strain was resistant to gentamicin and tobramycin but susceptible to amikacin. It showed also, decreased susceptibility to trimethoprim/sulfamides and rifampicin. The MIC with microbroth dilution technique was 512 µg/ml for norfloxacin, 256 µg/ml for ofloxacin and ciprofloxacin and 64 µg/ml for levofloxacin.

Detection of PMQR genes

The PMQR screening with PCR revealed the presence of *qnrD* in *M. morgani* only. None of the other studied Enterobacteria harboured the *qnrD* gene. The sequencing result showed the presence of *qnrD1* in the *M. morgani* isolate. *qnrA*, *qnrB*, *qnrS*, *qnrC*, *aac-Ib-cr* and *qepA* genes were not found in this strain.

Conjugation

The conjugative reaction fails and the *qnrD* gene couldn't be transferred to *E. coli* J53.

Topoisomerase mutations

Sequencing of PCR products of QRDRs in *gyrB* and *parC* showed two mutations in the QRDR of *gyrB* at 463 and 464 codons. The first one is an S463A substitution and the second is an S464Y substitution.

Mutation was found also in the QRDR of *parC* at codon 80. It was an S80I substitution.

Discussion

The Gram-negative *M. morgani* is found in the environment and as a part of the normal flora of humans. It has been considered as a rare cause of human infections. However, it is an important opportunistic pathogen in urinary tract, soft tissue infections and also in infections following surgery [12,13]. In this case of study, the old age of the patient, the bladder tumor and its resection and the probe introduction after the surgery are risk factors leading to the urinary tract infection with the opportunistic *M. morgani*.

This *M. morgani* strain was highly resistant to quinolones and *qnrD* was detected. *qnrD* was not found to be self transmissible and it could have spread either on a mobilizable plasmid or on a transferable structure integrated into a conjugative plasmid [6]. The transfer of quinolone resistance due to *qnrD* is always studied on transformants never on transconjugants. In the study of Hu et al., *Proteus mirabilis* harbouring *qnrD* failed to transfer quinolone resistance to *E.coli* 600 by conjugation [14]. The MIC of ciprofloxacin in *E.coli* DH10B cells increased in the transformants carrying the plasmid pBR322 with the cloned *qnrD* from 0,002 µg/ml to 0,06 µg/ml (by a factor 32) [2].

However, *qnrD* positive clinical strains described so far exhibited a high level of quinolone resistance with ciprofloxacin MIC ranging from 1 to higher than 32 µg/ml and they are likely to harbour other quinolone resistance mechanisms [14,15]. Topoisomerase mutations have not been always investigated in *qnrD* positive clinical strains in order to evaluate how both mechanisms interact. The *gyrA* and *parE* primers used to amplify the QRDRs for Enterobacteria were not available for *M. morgani*. The study of Mazzariol A et al. couldn't to amplify *gyrA* and *parC* genes in this bacteria and this is due to its unknown genome [15].

To date, none of *M. morgani* was described with *gyrB* mutations. In this report, we described for the first time, two new *gyrB* mutations in *M. morgani*. In gram-negative bacilli, *gyrB* mutations are rare in clinical strains and they have been described at positions 426, 431, 447, 463, 464 and 466. The Ser 463Ala mutation has been linked to *Klebsiella oxytoca* [11].

This is the first report of the previous mutation in *qnrD* harbouring *M. morgani*. The Ser 464 Phe mutation has been linked to quinolone resistant *S.enterica*, *P.aeruginosa* and *P.mirabilis* [16-18]. In this study, the *gyrB* Tyr464 mutation was different from those described previously.

For *M. morgani*, the first *ParC* mutation was reported in the Tunisian study of Mahrouki et al. [19]. It was an S80I mutation associated to the *qnrS* 1 gene.

Another *ParC* mutation (S80R) was also described previously in this *Proteaeae* [20]. In this study, we described the first S80I mutation in *qnrD*-positive *M. morgani*. This type of mutation was already observed in other Enterobacteria but never associated to the *qnrD* gene [21].

In summary, we have reported for the first time in Tunisia the rare *qnrD* in *M. morgani* and also found two new *gyrB* mutations (S463A and S464Y) and one *parC* substitution (S80I). In addition to *qnrD* gene, both mutations in *gyrB* and *ParC* could contribute to the high level of resistance to quinolones in this strain. The nosocomial infection caused by this bacterium invites further study of its epidemiologic evolution.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MNY carried the bacteria collection and typing antibiotic susceptibility, PCR screening, sequence blasting and writing the manuscript. IDR is project designer and co-supervisor of research work. QG Laboratory supervisor, she controlled the analysis, interpretation of data and controlled the manuscript. MM provided the necessary materials for collection, typing and doing the susceptibility tests. MA project designer and general supervisor of research group. MW General supervisor of research experiments and controlled the manuscript. All authors read and approved the final manuscript.

Author details

¹Faculty of Sciences, Bizerta Carthage University, Bizerta, Tunisia. ²Laboratory of Biologically Active Substances and Communicable Diseases (LR99ES27), Monastir University, Monastir, Tunisia. ³Institute of Antibiotics, Huashan Hospital, Fudan University, Shanghai 200040, China. ⁴Laboratory of Microbiology, Fattouma Bourguiba University Hospital, Monastir, Tunisia.

Received: 25 April 2014 Accepted: 13 July 2014

Published: 9 August 2014

References

1. Martínez-Martínez L, Pascual A, Jacoby GA: **Quinolone resistance from a transferable plasmid.** *Lancet* 1998, **351**:797–799.
2. Cavaco LM, Hasman H, Xia S, Aarestrup FM: **QnrD, a novel gene conferring transferable quinolone resistance in *Salmonella enterica* Serovar Kentucky and *bovismorbificans* strains of human origin.** *Antimicrob Agents Chemother* 2009, **58**:603–608.
3. Zhao J, Chen Z, Chen S, Deng Y, Liu Y, Tian W, Huang X, Wu C, Sun Y, Sun Y, Zeng Z, Liu J: **Prevalence and dissemination of *oqxAB* in *Escherichia coli* isolates from animals, farmworkers, and the environment.** *Antimicrob Agents Chemother* 2010, **54**:4219–4224.
4. Koncan R, Mazzariol A, Kocsis B, Fontana R, Lanzafame P, Rasmu M, Cornaglia M: **First detection in Europe of plasmid-mediated fluoroquinolone resistance *qnrD* determinant.** *Clin Microbiol Infect* 2010, **16**:S126.
5. Ogbolu DO, Daini OA, Ogunludun A, Alli AO, Webber MA: **High levels of multidrug resistance in clinical isolates of Gram-negative pathogens from Nigeria.** *Int J Antimicrob Agents* 2011, **37**:62–66.
6. Guillard T, Cambau E, Neuwirth C, Nenninger T, Mbadi A, Brasme L, Vernet-Garnier V, Bajolet O, Champas C: **Description of a 2,683-base-pair plasmid containing *qnrD* in two *Providencia rettgeri* isolates.** *Antimicrob Agents Chemother* 2011, **56**(1):565–568.
7. Cattoir V, Poirel L, Rotimi V, Soussy C, Nordmann P: **Multiplex PCR for detection of plasmid-mediated quinolone resistance *qnr* genes in ESBL-producing enterobacterial isolates.** *J Antimicrob Chemother* 2007, **60**:394–397.
8. Minghua W, Qinglan G, Xiaogang X, Xiaoying W, Xinyu Y, Shi W, Hooper D, Mingui W: **New plasmid-mediated Quinolone resistance gene, *qnrC*, found in a clinical isolate of *Proteus mirabilis*.** *Antimicrob Agents Chemother* 2009, **53**(5):1892–1897.
9. Park C, Robicsek A, Jacoby GA, Sahm D, Hooper D: **Prevalence in the United States of *aac(6)-Ib-cr* encoding a Ciprofloxacin-modifying enzyme.** *Antimicrob Agents Chemother* 2006, **50**(11):3953–3955.
10. Yamane K, Wachino J, Suzuki S, Arakawa Y: **Plasmid-mediated *qepA* gene among *Escherichia coli* clinical isolates from Japan.** *Antimicrob Agents Chemother* 2008, **52**(4):1564–1566.
11. Lascols C, Robert J, Cattoir V, Bébéar C, Cavallo J, Podglajen I, Ploy M, Bonnet R, Soussy C, Cambau E: **Type II topoisomerase mutations in clinical isolates of *Enterobacter cloacae* and other enterobacterial species harbouring the *qnrA* gene.** *Int J Antimicrob Agents* 2007, **29**:402–409.
12. Tsai MT, Yeh JT, Yang WC, Wu TH: **CAPD-related Peritonitis caused by *Morganella morgani*.** *PD* 2013, **33**:104–105.
13. Chen Y, Peng H, Shia W, Hsu F, Ken C, Tsao Y, Chen C, Liu C, Hsieh M, Chen H, Tang C, Ku T: **Whole-genome sequencing and identification of *Morganella morgani* KT pathogenicity-related genes.** *BMC Genomics* 2012, **13**(Suppl 7):S4.
14. Hu Y, Cai J, Zhang R, Zhou H, Sun Q, Chen G: **Emergence of *Proteus mirabilis* harboring *blaKPC-2* and *qnrD* in a Chinese Hospital.** *Antimicrob Agents Chemother* 2012, **56**(5):2278–2282.

15. Mazzariol A, Kocsis B, Koncan R, Kocsis E, Lanzafame P, Cornaglia G: **Description and plasmid characterization of *qnrD* determinants in *Proteus mirabilis* and *Morganella morganii*.** *Clin Microbiol Infect* 2012, **18**:E46.
16. Gensberg K, Jin YF, Piddock LJ: **A novel *gyrB* mutation in a fluoroquinolone-resistant clinical isolate of *Salmonella typhimurium*.** *FEMS Microbiol Lett* 1995, **132**:57–60.
17. Mounaimne H, Robert J, Jarlier V, Cambau E: **Type II topoisomerase mutations in ciprofloxacin-resistant strains of *Pseudomonas aeruginosa*.** *Antimicrob Agents Chemother* 1999, **43**:62–66.
18. Weigel LM, Anderson GJ, Tenover FC: **DNAgyrase and topoisomerase IV mutations associated with fluoroquinolone resistance in *Proteus mirabilis*.** *Antimicrob Agents Chemother* 2002, **46**:2582–2587.
19. Mahrouki S, Bourouis A, Chihi H, Ouertani R, Ferjani M, Moussa MB, Barguellil F, Belhadj O: **First characterisation of plasmid-mediated quinolone resistance-*qnrS1* co-expressed *bla*_{CTX-M-15} and *bla*_{DHA-1} genes in clinical strain of *Morganella morganii* recovered from a Tunisian Intensive Care Unit.** *Indian J Med Microbiol* 2012, **30**:437–441.
20. Andres P, Lucero C, Soler-Bistué A, Guerriero L, Albornoz E, Tran T, Zorreguieta A, PMQR-Group, Galas M, Corso A, Tolmasky M, Petroni A: **Differential distribution of plasmid-mediated Quinolone resistance genes in clinical enterobacteria with unusual phenotypes of Quinolone susceptibility from Argentina.** *Antimicrob Agents Chemother* 2013, **57**(6):2467–2475.
21. Lim S, Lim K, Lee H, Jung S, Kang M, Nam H: **Prevalence and molecular characterization of Fluoroquinolone-resistant *Escherichia coli* isolated from diarrheic cattle in Korea.** *J Vet Med Sci* 2010, **72**(5):611–614.

doi:10.1186/s12941-014-0034-4

Cite this article as: Nasri Yaiche *et al.*: Type II and type IV topoisomerase mutations in clinical isolates of *Morganella morganii* harbouring the *qnrD* gene. *Annals of Clinical Microbiology and Antimicrobials* 2014 **13**:34.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

