

SHORT REPORT Open Access

Use of vancomycin as a surrogate for dalbavancin in vitro susceptibility testing: results from the DISCOVER studies

Michael W Dunne^{1*}, Dan Sahm² and Sailaja Puttagunta¹

Abstract

Background: Dalbavancin is a lipoglycopepetide antibiotic with activity against gram positive pathogens recently approved for treatment of acute bacterial skin and skin structure infections. Pending the introduction of antimicrobial susceptibility tests, we examined the utility of vancomycin inhibitory concentrations to predict dalbavancin susceptibility in a panel of isolates obtained from phase 3 registration studies.

Findings: 99.6% of *Staphylococcus aureus* and 99.0% of beta-hemolytic streptococci which are susceptible to vancomycin will have an MIC at or below the US FDA susceptibility breakpoint for dalbavancin.

Conclusion: Vancomycin should be considered as a surrogate for *in vitro* dalbavancin susceptibility testing.

Keywords: Susceptibility testing, Vancomycin, Dalbavancin, Antimicrobial agents, Acute bacterial skin and skin structure infections

With the introduction of new antimicrobial agents comes a need for diagnostic tests available in the community which can confirm the *in vitro* susceptibility of target pathogens. Those diagnostic tests, however, require a significant amount of research and development and can only be approved once the susceptibility breakpoints for the agent are identified. Consequently, many antibacterial agents newly available for clinical use are not included in the automated testing systems commonly used in hospital settings.

Dalbavancin is a new lipoglycopeptide with activity against Gram-positive pathogens that has a long half-life allowing for infrequent dosing. Based on two phase 3 clinical trials [1] dalbavancin was recently approved in the United States for treatment of acute bacterial skin and skin structure infections (ABSSSI) dosed intravenously as 1000 mg followed one week later by 500 mg [2] (USPI). Antimicrobial susceptibility testing methodologies in broth have been established by the Clinical and Laboratory Standards Institute [3] (CLSI) however commercially available diagnostics are not presently available. In this

circumstance, the use of a surrogate agent to assess for *in vitro* susceptibility is well precedented and has significant utility pending the addition of the new antibacterial agent to an established diagnostic testing susceptibility platform [3,4].

Dalbavancin shares a similar mechanism of action with vancomycin, though providing a significant enhancement in potency, presumably a result of modification to one of the side chains as well as the addition of a lipid tail. The use of vancomycin as a surrogate for dalbavancin *in vitro* susceptibility has been previously proposed [5], and recently repeated [6] (R.N. Jones, D.J. Farrel, R. K. Flamm, submitted for publication), based on a large collection of isolates obtained from a hospital surveillance program. Recently, *in vitro* susceptibility data, derived from the DISCOVER clinical development program has also become available that allows for cross tabulation of mean inhibitory concentrations for these two agents.

Findings

Isolates derived from patients enrolled in the dalbavancin clinical development program for ABSSSI were tested for susceptibility against dalbavancin and vancomycin. Identification of isolates was confirmed by MALDI Biotyper

^{*} Correspondence: mdunne@duratatx.com

Durata Therapeutics, Inc., Branford, CT, Branford, CT, USA
Full list of author information is available at the end of the article



Table 1 CLSI methodology for broth microdilution

Testing conditions	Dalbavancin	Vancomycin Cation-adjusted Mueller Hinton Broth		
Medium	Cation-adjusted Mueller Hinton Broth supplemented with 0.002% (v/v) polysorbate 80			
Inoculum	Direct colony suspension, equivalent to a 0.5 McFarland standard	Direct colony suspension, equivalent to a 0.5 McFarland standard		
Incubation	35 +/- 2°C; ambient air; 24 hours	35 +/- 2°C; ambient air; 24 hours		
Solvent/diluent	DMSO	Water		

(Bruker, Bellerica, MA USA). Briefly, dalbavancin was diluted in DMSO and susceptibility testing was performed by broth microdilution in Cation-adjusted Mueller Hinton Broth supplemented with 0.002% (v/v) polysorbate 80, as described in CLSI M7, CLSI M100 and Eurofins Medinet SOP [7] (1-P-PR-PRO-9002355– Broth Microdilution MIC Testing with Frozen Panels). Quality control and interpretations of results were performed according with CLSI M100 with the reference strains *Staphylococcus aureus* ATCC 29213 and *Streptococcus pneumoniae* ATCC 49619. Vancomycin testing was performed in a similar manner, with differences as outlined in Table 1.

Of the 634 S. aureus isolates identified, the dalbavancin MIC₉₀ was 0.06 μ g/ml. Three isolates (0.4%) had an MIC greater than the FDA breakpoint of >0.12 μ g/ml. The MIC₉₀ of S. aureus for vancomycin was 1 μ g/ml and no isolate had an MIC greater the breakpoint of 2 μ g/ml. Cross tabulation of these susceptibility data confirmed that 99.6% of the isolates that were susceptible to vancomycin were also susceptible to dalbavancin (Table 2).

Of the 192 beta-hemolytic streptococci, which include *S. pyogenes* (87), *S. agalactiae* (36), *S. dysgalactiae* (4), *S. anginosus* group (49), Group C streptococci (9) and Group G streptococci (7), the dalbavancin MIC_{90} was 0.06 µg/ml. Two isolates had an MIC > 0.12 µg/ml. The two isolates (1%) that would be considered non-susceptible to dalbavancin were both susceptible to vancomycin. Cross tabulation of these susceptibility data confirmed that 99.0% of

Table 2 Cross tabulation of *S. aureus* isolates with mean inhibitory concentrations for vancomycin and dalbavancin

Vancomycin MIC (µg/ml)	Dalbavancin MIC (μg/ml)							
	0.008	0.015	0.03	0.06	0.12	0.25		
0.25		1						
0.5		3	142	191	5	1		
1	1	2	49	230	7	1		
2						1		

Table 3 Cross tabulation of Beta-hemolytic streptococcal isolates with mean inhibitory concentrations for vancomycin and dalbavancin

	Dalbavancin MIC (μg/ml)								
MIC (µg/ml)	<0.001	0.002	0.004	0.008	0.015	0.03	0.06	0.12	0.25
≤0.015	1	1			1				
0.03		1							
0.06				1					
0.12		1							
0.25		2	19	16	38	14	4	3	2
0.5		7	7	18	20	16	3	4	
1			1	4	3		1		

the isolates that were susceptible to vancomycin were also susceptible to dalbavancin (Table 3).

One patient treated with dalbavancin had an MIC that would be considered non-susceptible (MIC = 0.25 μ g/ml). This organism was positive for the Panton-Valentine leukocidin toxin and methicillin resistance and the patient was a clinical success at Day 3 and a clinical cure at Day 14. The other patient with a dalbavancin MIC to *S. aureus* of >0.12 μ g/ml (MIC = 0.25 μ g/ml) was treated with vancomycin and was a clinical success (Table 4).

Staphylococcal and beta-hemolytic streptococcal isolates were collected from two large clinical trials supporting the development of dalbavancin for treatment of ABSSSI. Based on recent epidemiologic surveys, the distribution of MICs in this collection is representative of isolates circulating in hospitals in the United States [8]. 99.6% of *S. aureus* and 99.0% of beta-hemolytic streptococci which are susceptible to vancomycin (MIC \leq 2 µg/ml) will have an MIC at or below the US FDA susceptibility breakpoint for dalbavancin.

While vancomycin susceptible isolates were also susceptible to dalbavancin, no data are available regarding the potential clinical outcome of patients treated with dalbavancin who have isolates with an MIC to

Table 4 Two patients with S. aureus isolates with a dalbavancin MIC of 0.25 $\mu g/ml$

Characteristic	Patient 1	Patient 2		
Treatment Group	Vancomycin/linezolid	Dalbavancin		
Age (years)	51	34		
Type of infection	Traumatic Wound Infection	Cellulitis		
Wound culture	MSSA	MRSA		
Vancomycin MIC	1	0.5		
PVL toxin	Negative	Positive		
mecA gene	Negative	Positive		
Clinical response at 48–72 hours	Responder	Responder		
Clinical outcome at Day 14	Success	Success		

vancomycin > 2 µg/ml. Anecdotally, a small number of these patients have been observed in clinical trials and have been successfully treated. While the FDA breakpoint is 0.12 µg/ml, based to a large degree on the clinical outcome data of a sufficient number of patients infected with isolates at that MIC, a preliminary assessment of the ECOFF calculations [9] for *S. aureus* suggests a breakpoint of 0.25 µg/mL, as this is the highest MIC of organisms lacking phenotypically expressed resistance [10]. A reassessment of the existing breakpoint will be enabled by analysis of more patients treated with dalbavancin who had organisms with an MIC > 0.12 µg/ml.

A strong correlation between vancomycin and dalbavancin *in vitro* susceptibility results was observed for both sets of isolates. While broth microdilution methodologies to guide performance of *in vitro* susceptibility testing have been published, these data support the proposal that vancomycin can be used as a surrogate for susceptibility testing of dalbavancin, pending the introduction of dalbavancin into established diagnostic susceptibility testing platforms.

Over 99.6% of *S. aureus* and 99.0% of beta-hemolytic streptococci which are susceptible to vancomycin (MIC \leq 2 µg/ml) will have an MIC at or below the US FDA susceptibility breakpoint for dalbavancin. Vancomycin should be considered for use as a surrogate for *in vitro* dalbavancin susceptibility testing.

Availability of supporting data

The data supporting the results of this study are included within this article.

Competing interests

Dr. Dunne is an employee of Actavis plc and was the Chief Medical Officer of Durata Therapeutics, Inc. Dr. Puttagunta is an employee of Actavis plc and was the VP of Development and Clinical Affairs at Durata Therapeutics, Inc. Dr. Sahm has nothing to disclose.

Authors' contributions

MWD and SP made substantial contributions to the acquisition of data and the analysis plan. MWD, SP and DS made substantial contributions to the analysis and interpretation of the data. MWD and SP were involved in drafting the manuscript. All authors have read and approved the final manuscript.

Acknowledgements

The authors appreciate the careful review of this manuscript by Sandra McCurdy and the administrative assistance of Shaina Barnes.

Author details

¹Durata Therapeutics, Inc., Branford, CT, Branford, CT, USA. ²International Health Care Associates, Inc., Washington, DC, USA.

Received: 15 December 2014 Accepted: 23 March 2015 Published online: 02 April 2015

References

 Boucher HW, Wilcox M, Talbot GH, Puttagunta S, Das AF, Dunne MW. Onceweekly dalbavancin versus daily conventional therapy for skin infection. NEJM. 2014;370(23):2169–78.

- DalvanceTM Package Insert 2014. DurataTherapeutics Intl., Chicago, IL, USA. Available at http://www.accessdata.fda.gov/drugsatfda_docs/label/2014/ 021883s000lbl.pdf. Accessed July 2014
- Clinical and Laboratory Standards Institute. M100-S24. Performance standards for antimicrobial susceptibility testing: 24th informational supplement. Wayne, PA: CLSI; 2014.
- Friedland IR, Isaacs R, Mixson LA, Motyl M, Woods GL. Use of surrogate antimicrobial agents to predict susceptibility to ertapenem. Diagn Microbiol Infect Dis. 2002;43:61–4.
- Jones RN, Sader HS, Fritsche TR, Hogan PA, Sheehan DJ. Selection of a surrogate agent (vancomycin or teicoplanin) for initial susceptibility testing of dalbavancin: results from an international antimicrobial surveillance program. J Clin Microbiol. 2006;44:2622–5.
- Jones RN, Farrell DJ, Flamm RK, Sader HS, Dunne MW, Mendes RE. Surrogate Analysis of Vancomycin to Predict Susceptible Categorization of Dalbavancin. Diagn Microbiol Infect Dis. 2015 Published online Feb 7. doi:10.1016/j.diagmicrobio.2015.01.017.
- CLSI M100-S24. Performance Standards for Antimicrobial Susceptibility Testing; Twenty-Fourth Informational Supplement. 2014, 34: 1.
- Jones RN, Sader HS, Flamm RK. Update of dalbavancin spectrum and potency in the USA: report from the SENTRY Antimicrobial. Surveillance Program (2011). 2013;75:304–7.
- Kronvall G, Kahlmeter G, Myhre E, Galas MF. A new method for normalized interpretation of antimicrobial resistance from disk test results for comparative purposes. Clin Microbiol Infect. 2003;9:120–32.
- Kahlmeter G, EUCAST. ECOFF presentation to clinical laboratory and standards institute meeting. Tampa, FL USA: EUCAST; 2013.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at www.biomedcentral.com/submit

